

Helicoidal transfer matrix model for inhomogeneous DNA melting

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An inhomogeneous helicoidal nearest-neighbor model with continuous degrees of freedom is shown to predict the same DNA melting properties as traditional long-range Ising models, for free DNA molecules in solution, as well as superhelically stressed DNA with a fixed linking number constraint. Without loss of accuracy, the continuous degrees of freedom can be discretized using a minimal number of discretization points, yielding an effective transfer matrix model of modest dimension ($d=36$). The resulting algorithms to compute DNA melting profiles are both simple and efficient.

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I. INTRODUCTION

The computation of the thermal stability and statistical physics of nucleic acids is a classical problem going back to the 1960's. The standard model to describe the untwisting and separation of both strands of a free DNA double-helix in solution is the Poland-Scheraga helix-coil model, where each base pair can be in two possible states, helix (closed) or coil (open) [1,2]. The addition of entropy weights to a basic Ising model, counting the number of possible configurations of open loops, induces an effective long range interaction between base pairs which is essential for correctly obtaining the helix specific opening probabilities. The most widely used algorithm for computing the opening probabilities is the recursion relation method of Poland [3]. Incorporating the Fixman-Freire approximation [4] for the loop entropy factor reduces the computational complexity from $O(N^2)$ to $O(N)$ in the sequence length N . With the availability of fully sequenced genomes, the study of DNA melting or denaturation has become an active field of research again with recent results relating the physics of denaturation to the biology of genomes [5,6], reparametrizing the original loop entropy weights [7], speeding up the Poland-Fixman-Freire algorithm for whole genome sequences [8], and generalizing the model to describe hybridization with mismatches of unequal length sequences [9]. The traditional physics approach to compute statistical mechanical probabilities by transfer matrix multiplication [10,11] has also recently been revisited by Poland [12]. While this last algorithm offers no improvement in computational complexity [using matrix sparsity it is $O(N^2)$], it is very simple and straightforward to implement.

In vivo DNA strand separation involves interactions with other molecules which impose superhelical stresses on the DNA molecule. This is modeled by Benham's statistical mechanical model for stress induced duplex destabilization (SIDD) [13], which also is a helix-coil model with Ising degrees of freedom. It has a long range base pair interaction arising through superhelical constraints (no loop entropy fac-

tors are added), and opening probabilities are known to correlate very well with regions important for transcriptional regulation [14,15]. An exact solution of the model is $O(N^2)$ but an accelerated algorithm using an energy cutoff reduces this to $O(N)$, such that SIDD properties can be computed for whole genome sequences as well [16,17].

In parallel with the helix-coil models, a distinct class of statistical mechanical models for DNA melting has been developed starting from a physically more realistic description of a base pair as an entity which has a continuum of intermediate states in between helix or coil. These models are all based on the Peyrard-Bishop model [18] which consists of a nonlinear particle lattice with one real degree of freedom per base pair describing the stretching of the hydrogen bonds between the bases. Nonlinearity and cooperativity in such a model arises already with a nearest-neighbor interaction, no long-range interaction is needed [19]. Subsequent improvements to the model include replacing the harmonic stacking energy by an anharmonic stacking energy [20], and introducing an additional angular degree of freedom per base pair to model the helicoidal structure of DNA [21–23]. In the latter model, separation of the two strands is coupled to the untwisting of the double helix. The effect of sequence inhomogeneity on the melting transition in the Peyrard-Bishop model with harmonic and anharmonic stacking has been studied for random sequences [24] and for periodic sequences [25].

Recent experimental developments (see [26] for a review) have made it possible to manipulate single polymeric molecules directly and thus offer access to a whole range of DNA properties other than the melting phenomenon. These elasticity experiments too can be accurately modeled by yet another type of statistical mechanical models consisting of a double-helix with nonopening base pairs connected by flexible, folding backbones [27,28]. In this paper however, we will be concerned with the melting transition only and the connection between the continuous particle-lattice models and the discrete helix-coil models.

Unlike the helix-coil models, which have seen many applications to real biological sequences, the particle-lattice models are mostly used to obtain a more fundamental, sequence independent, physical understanding of the DNA

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melting phenomenon, such as the order of the phase transition, the existence of nonlinear “bubble” excitations, etc., (see [19] for a recent review paper). Moreover, although both types of models have been validated against (different) experiments, very little is known about how they relate to one another and whether they are in some sense equivalent. Here, we attempt to close the gap between both kinds of models. We study an inhomogeneous particle-lattice model based on the Barbi-Cocco-Peyrard helicoidal model [21] and compute its melting properties for some standard example sequences both under free conditions and with superhelical stresses.

The thermally induced melting of free DNA is obtained as the formally very simple transfer integral equilibrium solution of the helicoidal model, yet computation of the melting properties is a challenge in itself as, e.g., a computation of the partition function involves $O(N)$ numerical integrations over an infinite integration domain. However, Zhang *et al.* [25] already observed that for the Dauxois-Peyrard-Bishop model [20], the numerical integrations can be carried out using a very limited number of discretization points; a dimension as small as $d=70$ gave very accurate results compared to much higher dimensions ($d=800$ and more), and by allowing an error of order 10^{-6} with respect to the exact results, the dimension could be further reduced to $d \approx 40$. For the helicoidal model, we have found a value of $d=36$ to be the minimal discretization dimension. This effectively reduces the particle-lattice model to a nearest-neighbor generalized Ising model, offering the possibility to develop a very simple and very fast algorithm to compute DNA melting probabilities. We propose such an algorithm which, moreover, is numerically stable for arbitrary sequence lengths, avoiding underflow problems related to the extensivity of the free energy (i.e., the exponential vanishing of the partition function for diverging sequence length). The algorithm is as simple as Poland’s matrix algorithm [12] and as fast as any of the fastest helix-coil algorithms discussed above. Extension of the algorithm to compute correlations between different base pairs, loop opening probabilities, higher order moments for base pair opening, etc., is trivial and straightforward.

Stress induced DNA melting is modeled by imposing a fixed linking number constraint on the DNA strands, which leads to a coupling of all angular degrees of freedom in the model. However, the linking number is thermodynamically conjugated to an external torque variable applied at both ends of the molecule. The model with external torque can be solved by the above transfer matrix algorithm, and although there is no equivalence of ensembles, the fixed linking number solution can be obtained by a complex integration over the torque variable. The numerical solution is $O(MN)$, where M is a constant independent of N determined by the desired accuracy of the torque integration, a situation similar to the analysis of stress induced DNA melting using Benham’s SIDD model [17].

II. THE MODEL AND ITS EQUILIBRIUM SOLUTION

We consider the helicoidal model introduced by Barbi, Cocco, and Peyrard [21]. Unlike the original homogeneous

model, the various energy parameters will be explicitly sequence dependent. A DNA sequence is a string of N letters $\{A, C, G, T\}$, which for convenience we translate (alphabetically) into a numerical sequence $(s_n)_n$ taking values in $\{1, 2, 3, 4\}$. Each base pair in the model has two degrees of freedom, a radial variable r , related to the opening of the hydrogen bonds, and an angular variable ϕ , related to the twisting of the base pair and responsible for the three-dimensional structure of the DNA molecule. Successive angles are restricted to $\phi_{n+1} - \phi_n \in [0, \pi]$ to enforce helical geometry. Alternatively, we can associate a radial variable r to the sites of the lattice, and an angular variable $\theta \in [0, \pi]$ to the bonds of the lattice ($\theta_n = \phi_{n+1} - \phi_n$).

The potential energy is given by

$$V = \sum_{n=1}^N D_{s_n} (e^{-a_{s_n}(r_n - r_0)} - 1)^2 + \sum_{n=1}^{N-1} K_{s_n s_{n+1}} (r_{n+1} - r_n)^2 e^{-\alpha(r_n + r_{n+1} - 2r_0)} + \sum_{n=1}^{N-1} E_{s_n s_{n+1}} (\ell_{n,n+1} - \ell_{s_n s_{n+1}}^{(0)})^2 - \Gamma \sum_{n=1}^{N-1} \theta_n. \quad (1)$$

The first term is the Morse potential modeling the hydrogen bonds between the two nucleotides in a base pair [18], the second term is the anharmonic stacking interaction between successive base pairs [20], the third term is a harmonic twist energy allowing fluctuations of the length $\ell_{n,n+1}$ between successive nucleotides on the same DNA strand [21] (see also the Appendix), and the last term, which can be written as $-\Gamma(\phi_N - \phi_1)$, is the external torque or superhelical twist. $\Gamma > 0$ overtwists the DNA molecule, inhibiting denaturation of the two strands, $\Gamma < 0$ causes undertwisting and enhances denaturation [23].

We note here that the present model is only suitable for negative or small positive torque Γ . Indeed, unwinding leads to denaturation and this is well described by the potential energy (1), but severe overwinding leads to DNA forms with exposed bases and the backbone winding at the center [26]. Such transition can obviously not be part of the present model.

A variety of boundary conditions (b.c.) can be considered for the radial variable r , such as free b.c., fixed b.c., or periodic b.c., with minor modifications to the numerical solution of the model. For the angular variable ϕ we consider two distinct situations. The first is to set $\phi_1 = 0$ and have no constraint on ϕ_N , corresponding to free b.c. for the variables θ , and describing the situation in some single molecule experiments [26]. The second situation, modeling superhelical stresses, is to set a fixed linking number constraint $\phi_N - \phi_1 = \sum_n \theta_n = \alpha N$, $\alpha \in [0, (N-1)\pi/N]$, which contains periodic b.c. in ϕ as the special case $\alpha = 2\pi n/N$, $n = 1, 2, 3, \dots$. The torque Γ and the total twist $\sum_n \theta_n$ are thermodynamically conjugated variables, yet as we are explicitly working with a finite-size system, there is no equivalence of ensembles and both situations lead to different melting properties. We will

refer to the first situation as the ‘‘torque ensemble’’ and the second as the ‘‘linking number ensemble.’’

The choice of the energetic parameters is a difficult one and unlike for the helix-coil models, no well established set of parameters exists, especially with respect to the base pair dependence of the different energy terms. Morse potential constants for weakly bonded $A-T$ vs strongly bonded $C-G$ base pairs have been determined by comparison of the Dauxois-Peyrard-Bishop model with denaturation experiments [29]. For the other parameters, we follow the classification of El Hassan and Calladine [30]. More precisely we take $K_{s,t}$ inversely proportional to the slide variance of the step (s,t) , and $E_{s,t}$ inversely proportional to the twist variance ([30] Table II). To obtain explicit values, we first adapt the relative strength of the energy parameters such that their order of magnitude agrees with the parameters used in [23]. In that case we obtain the correct transition temperature interval, but a less perfect differential melting map (a melting map gives for each base pair the temperature at which it transforms from closed to open, see [31] and Sec. III A). By increasing the relative strength of the twist energy, the transition interval is widened, but the melting map becomes exact. To compute opening probabilities and melting maps, and identify stable vs unstable regions, this last set of parameters is more adequate. A more detailed comparison with an experiment will be needed to find the parameters which best fit the physical melting transition, but we do not pursue this further in this paper. All the explicit numerical values we use are given in the Appendix. To conclude, we mention that in the torque ensemble, to first order, sequence specificity in the melting process comes from the base pair specific Morse potentials, but inhomogeneity in the stacking and twist energies has second order effects which are nonetheless important for a detailed identification of the different melting domains. In the linking number ensemble, the coupling of all angular degrees of freedom leads to more complicated sequence specific melting behavior.

A. Equilibrium solution in the torque ensemble

Since we are not interested in velocity dependent quantities, the kinetic energy terms can be integrated directly in the partition function, which becomes, up to a multiplicative constant and with free b.c.,

$$Z = \int dr_1 \cdots \int dr_N \int d\theta_1 \cdots \int d\theta_{N-1} r_1 \cdots r_N e^{-\beta V}. \quad (2)$$

The θ -integrals factorize, and

$$Z = \int dr_1 \cdots \int dr_N T^{(1)}(r_1, r_2) \cdots T^{(N-1)}(r_{N-1}, r_N),$$

where for $n=1, \dots, N-2$,

$$T^{(n)}(r, r') = r e^{-\beta V_m^{(n)}(r)} e^{-\beta V_s^{(n)}(r, r')} \times \int_{-1}^1 \frac{dx}{\sqrt{1-x^2}} e^{-\beta V_t^{(n)}(r, r', x)} e^{\beta \Gamma \cos(x)},$$

and

$$T^{(N-1)}(r, r') = r r' e^{-\beta [V_m^{(N-1)}(r) + V_m^{(N)}(r')]} e^{-\beta V_s^{(N-1)}(r, r')} \times \int_{-1}^1 \frac{dx}{\sqrt{1-x^2}} e^{-\beta V_t^{(N-1)}(r, r', x)} e^{\beta \Gamma \cos(x)}.$$

$V_m^{(n)}$, $V_s^{(n)}$, and $V_t^{(n)}$ denote, respectively, the Morse, stacking, and twist energy terms. As we will not need spectra of transfer integral operators, there is no need for symmetrizing these kernels.

In order to compute expectation values of the form $\langle f(r_n) g(\cos \theta_n) \rangle$ for the suitable test functions f and g , we need additional transfer integral operators

$$T_{f,g}^{(n)}(r, r') = r f(r) e^{-\beta V_m^{(n)}(r)} e^{-\beta V_s^{(n)}(r, r')} \times \int_{-1}^1 \frac{dx}{\sqrt{1-x^2}} g(x) e^{-\beta V_t^{(n)}(r, r', x)} e^{\beta \Gamma \cos(x)},$$

with appropriate modifications for the right-most sites $N-1$ and N .

Since strand separation and untwisting are directly correlated by the twist energy term, it is often sufficient to consider the case $g \equiv 1$, such that we get the simpler kernels

$$T_f^{(n)}(r, r') = f(r) T^{(n)}(r, r'), \quad (3)$$

$$T_f^{(N)}(r, r') = T^{(N-1)}(r, r') f(r'). \quad (4)$$

To solve the model numerically, we replace the transfer integral operators by finite size transfer matrices. The most efficient way for doing this is approximating the integrals in the partition function by finite sums using Gaussian quadratures [32]. For the angular x -integrals, this is straightforward as they already contain the right weight function for Gauss-Chebyshev integration. For the radial r -integrals, we first restrict the infinite integration domain to a finite interval $[a, b]$, then apply the Gauss-Legendre integration. Let z_j , $j=1, \dots, M_C$, be the zeros of the M_C th Chebyshev polynomial, all having equal weight π/M_C . Let z'_j , $j=1, \dots, M_L$, be the zeros of the M_L th Legendre polynomial, $\xi_j = 1/2(b-a)z'_j + 1/2(b+a)$ the zeros transformed to the interval $[a, b]$, and w_j the associated weights [32].

We obtain the transfer matrix approximation to the partition function,

$$Z = \sum_{i,j} (\hat{T}^{(1)} \cdots \hat{T}^{(N-1)})_{ij} = \langle v | \hat{T}^{(1)} \cdots \hat{T}^{(N-1)} | v \rangle,$$

where $v = (11 \cdots 1)$, $|\cdot\rangle$ and $\langle \cdot|$ are the familiar Dirac column, respectively, row vector notation, and $\hat{T}^{(n)}$ are the $M_L \times M_L$ transfer matrices defined by

$$\hat{T}_{ij}^{(n)} = w_i \xi_i e^{-\beta V_m^{(n)}(\xi_i)} e^{-\beta V_s^{(n)}(\xi_i, \xi_j)} \times \frac{\pi}{M_C} \sum_{k=1}^{M_C} e^{-\beta V_t^{(n)}(\xi_i, \xi_j, z_k)} e^{\beta \Gamma \cos(z_k)}$$

$$\hat{T}_{ij}^{(N-1)} = w_i w_j \xi_i \xi_j e^{-\beta[V_m^{(N-1)}(\xi_i) + V_m^{(N)}(\xi_j)]} e^{-\beta V_s^{(n)}(\xi_i, \xi_j)} \\ \times \frac{\pi}{M_C} \sum_{M_C k=1}^{M_C} e^{-\beta V_i^{(n)}(\xi_i, \xi_j, z_k)} e^{\beta \Gamma \text{acos}(z_k)}.$$

Likewise matrices $\hat{T}_{f,g}^{(n)}$ are defined as finite approximations to the corresponding kernels.

Different boundary conditions in the radial variable can be easily accommodated by changing the vector v : for fixed b.c. $r_1=r_N=\xi_j$, $v_i=\delta_{ij}$, for closed, resp. open b.c., $v_i=I(\xi_i \leq 12)$, resp. $v_i=I(\xi_i > 12)$, and for periodic b.c. the inner product $\langle v | \cdot | v \rangle$ is replaced by the trace $\text{Tr}(\cdot)$. Here we follow the convention that a base pair is ‘‘open’’ if $r-r_0 > 2\text{\AA}$, and I denotes the indicator function, $I(A)=1$ if the condition A is true.

Defining left and right matrix products

$$M_L^{(n)} = \hat{T}^{(1)} \dots \hat{T}^{(n)}, \quad M_R^{(n)} = \hat{T}^{(n)} \dots \hat{T}^{(N-1)}$$

for $n=1, \dots, N-1$, and $M_L^{(0)}=M_R^{(N)}=1$, we obtain

$$\langle f(r_n)g(\cos \theta_n) \rangle = \frac{\langle v | M_L^{(n-1)} \hat{T}_{f,g}^{(n)} M_R^{(n+1)} | v \rangle}{Z}$$

$$\langle f(r_N) \rangle = \frac{\langle v | M_L^{(N-2)} \hat{T}_f^{(N)} | v \rangle}{Z}.$$

The different transfer matrices $\hat{T}^{(n)}$ for $n=1, \dots, N-2$ choose between 16 different matrices, one for each nucleotide step-type. These matrices, together with one matrix $\hat{T}^{(N-1)}$ for the final bond, are computed first and stored on disk. For a given sequence we then compute and store the left and right matrix products. For a given pair (f, g) we compute again first the 16 possible matrices, $\hat{T}_{f,g}^{(n)}$, and the two matrices, $\hat{T}_{f,g}^{(N-1)}$ and $\hat{T}_f^{(N)}$. With these matrices, we can then compute, e.g., a profile $n \mapsto \langle f(r_n)g(\cos \theta_n) \rangle$ by the above formulas. By the simplicity of the transfer matrix formalism, the computational complexity of this procedure clearly increases only linearly with N .

However, even for sequences of moderate length (a few kilobase pair with double precision calculations), the left and right matrix products have such small entries, that they consist of round-off error only, and the computations become meaningless. This is a common problem due to the extensivity of the free energy. To make this computation work for sequences of arbitrary length, we define normalized left and right vectors;

$$\langle w_L^{(n)} | = \frac{\langle w_L^{(n-1)} | \hat{T}^{(n)} \rangle}{\| \langle w_L^{(n-1)} | \hat{T}^{(n)} \rangle \|}, \quad | w_R^{(n)} \rangle = \frac{\hat{T}^{(n)} | w_R^{(n+1)} \rangle}{\| \hat{T}^{(n)} | w_R^{(n+1)} \rangle \|},$$

with $w_L^{(0)}=w_R^{(N)}=v/\|v\|$, and while inductively creating these vectors we store

$$c_n = \| \hat{T}^{(n)} | w_R^{(n+1)} \rangle \|.$$

A short calculation reveals that

$$\langle f(r_n)g(\cos \theta_n) \rangle = \frac{\langle w_L^{(n-1)} | \hat{T}_{f,g}^{(n)} | w_R^{(n+1)} \rangle}{c_n \langle w_L^{(n-1)} | w_R^{(n)} \rangle}$$

$$\langle f(r_N) \rangle = \frac{\langle w_L^{(N-2)} | \hat{T}_f^{(N)} | w_R^{(N)} \rangle}{c_{N-1} \langle w_L^{(N-2)} | w_R^{(N-1)} \rangle},$$

involving only normalized vectors, sequence length independent (f, g) matrices, and the constants c_n , which are formed by sequence length independent matrices acting on normalized vectors.

If $g \equiv 1$, transfer matrices are of the form (3) and (4), and the formulas are even simpler. Denote by D_f the multiplication operator with the function f and by \hat{D}_f its diagonal matrix discretization. We get

$$\langle f(r_n) \rangle = \frac{\langle w_L^{(n-1)} | \hat{D}_f | w_R^{(n)} \rangle}{\langle w_L^{(n-1)} | w_R^{(n)} \rangle} \quad (5)$$

$$\langle f(r_N) \rangle = \frac{\langle w_L^{(N-2)} | \hat{T}^{(N-1)} \hat{D}_f | w_R^{(N)} \rangle}{\langle w_L^{(N-2)} | w_R^{(N-1)} \rangle}. \quad (6)$$

This method can be easily extended to compute higher moments. For example, to compute $\langle f(r_n)f(r_m) \rangle$ for fixed n and all m , we define $\hat{T}^{(n)'} = \hat{T}_f^{(n)}$ and $\hat{T}^{(l)'} = \hat{T}^{(l)}$ for $l \neq n$. Writing $\langle \cdot \rangle'$ to denote expectation with respect to these transfer matrices, we have, for functions $f > 0$,

$$\langle f(r_n)f(r_m) \rangle = \langle f(r_n) \rangle \langle f(r_m) \rangle'. \quad (7)$$

The practical applicability of the method clearly relies on the grid size values M_L and M_C , which were determined as follows. First we started from the value $M_L=70$, which according to Zhang *et al.* [25] gives exact results for the Peyrard-Bishop model. For this value, the upper limit of the integration domain has to be set to $b=40$, larger values of b require larger M_L [25]. The lower limit a can be put equal to 9.7 as the Morse potential can be considered infinite for smaller values. We determined a value $M_C=35$ to give accurate results in comparison with the MELTSIM program [31]. Like in the Peyrard-Bishop model [25], we then found that M_L could be further decreased with negligible error, to a value of 36. Further reducing the dimension leads to a dramatic change, where suddenly all the interesting transitional behavior is lost, the chain is either completely open, or completely closed. After M_L was minimized, we decreased M_C . Around $M_C=20$, we loose again all interesting behavior, but the transition is less sharp in this case. We settled on $M_C=24$.

The computational method presented so far works well up to a certain sequence length, where memory becomes the bottleneck instead of the CPU speed (around 10^6 bp on a typical PC). To treat even longer sequences, the sequence is divided into a number of smaller overlapping subsequences and the probability profiles of those are combined to obtain the full-length profile. This is a standard procedure [12,17], which, however, is much simpler in a nearest-neighbor model than in the long-range helix-coil models. More precisely, assume we cut the sequence of N base pairs into N/N_0

subsequences of length N_0 . To correct for the artificial boundaries thus introduced, we compute the opening probabilities for an interval $[lN_0-d, (l+1)N_0+d]$ but only keep the values for the interval $[lN_0, (l+1)N_0]$. If d is much larger than the typical *correlation* length, this gives the exact opening probability for the full sequence. In the helix-coil model, such a cut is never exact because d is always smaller than the *interaction* length. In Sec. III A, we will see that at typical values of T and Γ (i.e., values differentiating between stable and unstable regions), the correlation length is typically rather short, a few 100 base pairs at most. Therefore, a window size of length $N_0=10^5$ and overlap $2d$ between 10^3 and 10^4 leads to an exact algorithm for long sequences whose speed is only mildly affected by the windowing procedure.

B. Equilibrium solution in the linking number ensemble

The partition function in the linking number ensemble is again given by an integral of the form (2), but the angular integrals are now restricted to the subspace of $[0, \pi]^{N-1}$ for which the linking number or total twist satisfies

$$\frac{1}{N} \sum_{n=1}^{N-1} \theta_n = \alpha$$

for some fixed $\alpha \in [0, (N-1)\pi/N]$. Very often, instead of α , the superhelical density σ is specified,

$$\sigma = \frac{Lk - Lk_0}{Lk_0},$$

where $Lk = (2\pi)^{-1} \sum_n \theta_n$ is the linking number, and $Lk_0 = (2\pi)^{-1} \sum_n \theta_{n,n+1}^{(0)}$ is the ground state, zero torque linking number. As we remarked in Sec. II, our model is only suitable for negative or small positive superhelicity, and for definiteness we will consider in this section only negative torque Γ , corresponding to the negative superhelicity σ .

The situation is completely analogous to the standard statistical mechanics situation of canonical and grand-canonical ensembles; α plays the role of the “density,” Γ the role of a “chemical potential,” and by changing in Eq. (2) the angular integration variables to $\theta_1, \dots, \theta_{N-2}$, $\lambda = \sum_n \theta_n$, we see that the grand-canonical (torque ensemble) and canonical (linking number ensemble) partition functions are related by a Laplace transform

$$Z_{lq}(\Gamma) = \int_0^\infty d\lambda e^{\beta\Gamma\lambda} Z_{lk}\left(\frac{\lambda}{N}\right),$$

where it is understood that $Z_{lk}(\alpha) = 0$ for $\alpha > (N-1)\pi/N$.

By standard inverse Laplace transform techniques, the linking number partition function can be obtained from the torque partition function by a contour integration in the complex plane

$$Z_{lk}(\alpha) = \beta \int_{\Gamma-i\infty}^{\Gamma+i\infty} \frac{dz}{2\pi i} e^{-\beta z N \alpha} Z_{lq}(z) = \beta \int_{\Gamma-i\infty}^{\Gamma+i\infty} \frac{dz}{2\pi i} e^{-\beta N [z\alpha + F_{lq}(z)]}, \quad (8)$$

where the integral is carried out on a line parallel to the imaginary axis, with $\Gamma < 0$, and $F_{lq}(z)$ is the free energy in the torque ensemble.

Standard statistical mechanics proceeds by choosing a line which crosses the real axis at a critical point of the harmonic function $z\alpha + F_{lq}(z)$. This point is a saddle-point and the contour is a path of steepest descent, such that for large βN , the integrand in Eq. (12) is significantly different from zero in a small interval near the real axis only. Since we are interested in inhomogeneous, finite sequences, we do not consider the question of equivalence of ensembles in the thermodynamic limit.

More precisely, for a given $\sigma < 0$ and corresponding α , the function,

$$\Gamma < 0 \mapsto \alpha\Gamma + F_{lq}(\Gamma),$$

attains a maximum at some value $\Gamma_0 < 0$, namely the value Γ_0 for which $\langle \sum_n \theta_n \rangle_{lq, \Gamma_0} = N\alpha$, where $\langle \cdot \rangle_{lq, \Gamma_0}$ denotes expectation in the torque ensemble. The line passing through Γ_0 is chosen as the integration contour, and we can write

$$Z_{lk}(\alpha) = \beta e^{-\beta N \Gamma_0 \alpha} e^{-\beta N F_{lq}(\Gamma_0)} \times \int_{-\infty}^{\infty} \frac{d\omega}{2\pi} e^{-i\beta\omega N \alpha} e^{-\beta N [F_{lq}(\Gamma_0+i\omega) - F_{lq}(\Gamma_0)]}. \quad (9)$$

Because of the large parameter βN , the function,

$$\omega \mapsto |e^{-\beta N [F_{lq}(\Gamma_0+i\omega) - F_{lq}(\Gamma_0)]}|,$$

is tightly concentrated around $\omega=0$, and the integral can be restricted to a small interval $[-\epsilon_N, \epsilon_N]$. It is important to remark that to apply the standard stationary phase expansion, ϵ_N would have to be much smaller than $(\beta N)^{-1/2}$, a condition which is typically *not* fulfilled here. An efficient method to numerically compute the remaining integral consists of computing the integrand at a number of points and find a cubic splines interpolation which can be readily integrated.

The algorithm to compute expectation values proceeds as follows. Like in the previous section, let (f, g) be single-site test functions and denote by $Z^{(n)}$ the partition functions obtained by substituting at position n the transfer matrix $\hat{T}_{f,g}^{(n)}$ for $\hat{T}^{(n)}$. Further denote by $p_{lq}^{(n)}(\Gamma) = \langle f(r_n) g(\cos \theta_n) \rangle_{lq, \Gamma}$ the expectation value at torque Γ and analogously $p_{lk}^{(n)}(\alpha)$. Recalling that $p_{lq}^{(n)}(\Gamma) = \exp(-\beta N [F_{lq}^{(n)}(\Gamma) - F_{lq}(\Gamma)])$, we find

$$p_{lk}^{(n)}(\alpha) = p_{lq}^{(n)}(\Gamma_0) \times \frac{\int d\omega p_{lq}^{(n)}(\Gamma_0 + i\omega) e^{-i\beta N \alpha \omega} e^{-\beta N [F_{lq}(\Gamma_0+i\omega) - F_{lq}(\Gamma_0)]}}{\int d\omega e^{-i\beta N \alpha \omega} e^{-\beta N [F_{lq}(\Gamma_0+i\omega) - F_{lq}(\Gamma_0)]}}. \quad (10)$$

Since the lhs of this equation is obviously real, we take the

real parts of the integrands before numerically computing the integral. The torque expectation values $p_{iq}^{(n)}(\Gamma)$ are evaluated using the efficient algorithm of Sec. II A, which can be easily extended to also return the free energy,

$$\begin{aligned} -\beta N F_{iq}(\Gamma) &= \ln \langle v | \hat{T}^{(1)} \dots \hat{T}^{(N-1)} | v \rangle \\ &= \ln \langle v | \hat{T}^{(1)} | v \rangle + \sum_{n=2}^{N-1} \ln \frac{\langle v | \hat{T}^{(1)} \dots \hat{T}^{(n)} | v \rangle}{\langle v | \hat{T}^{(1)} \dots \hat{T}^{(n-1)} | v \rangle} \\ &= \sum_{n=1}^{N-1} \ln \frac{\langle w_L^{(n-1)} | \hat{T}^{(n)} | w_R^{(n)} \rangle}{\langle w_L^{(n-1)} | w_R^{(n)} \rangle}. \end{aligned}$$

Hence the algorithm to compute expectation values for all n in (10) is still $O(N)$, but M times slower than in the torque ensemble, where M only depends on the number of discretization points chosen to compute the ω -integrals.

To prove equivalence of the ensembles between the torque and linking number ensemble (for the given test functions) in the thermodynamic limit for homogeneous sequences, we would have to show that the fraction of the integrals in Eq. (10) tends to 1. For finite, inhomogeneous sequences, the nontriviality of this fraction causes a nonlinear coupling between base pairs that will be illustrated in Sec. III B.

III. EXAMPLE RESULTS

A. Torque ensemble

For easy comparison with the Poland-Scheraga helix-coil model, we show example results for the PN/MCS13 sequence ($N=4608$) which is the main example of [31]. This sequence is the pBR322 sequence [36] with an insert $[AAGTTGAACAAAAR]_{17}AAGTTGA$ at position 972 [33] ($[\dots]_x$ means $[\dots]$ x times repeated). The conclusions drawn here are equally valid for all other sequences we tested.

In the Peyrard-Bishop and related models a base pair is said to be denatured when it is stretched more than 2 \AA away from its equilibrium length of 10 \AA , hence the probability of denaturation is given by

$$p_n = \langle h(r_n - 12) \rangle, \quad (11)$$

where $h(r)$ is the Heaviside function, $h=1$ for $r \geq 0$ and 0 otherwise. Notice that we only need the simpler formulas (5) and (6) to compute melting profiles $n \mapsto p_n$.

Figure 1 shows the melting profile for the PN/MCS13 sequence at typical *in vivo* temperature $T=310 \text{ K}$. The torque value $\Gamma = -0.042 \text{ eV/rad}$ is chosen to give a good delineation of unstable regions ($p_n \approx 1$). Decreasing Γ increases the number of open base pairs, and increasing Γ has the opposite effect. On the basis of this melting profile we identify three unstable regions, the first one around position 1000 corresponding to the *AT*-rich insert in the pBR322 sequence, and the other two with maximums at position 3489 and at position 4423.

For whole genome sequences of several million base pairs, the opening probability is computed by dividing the sequence in shorter overlapping subsequences (see the end of

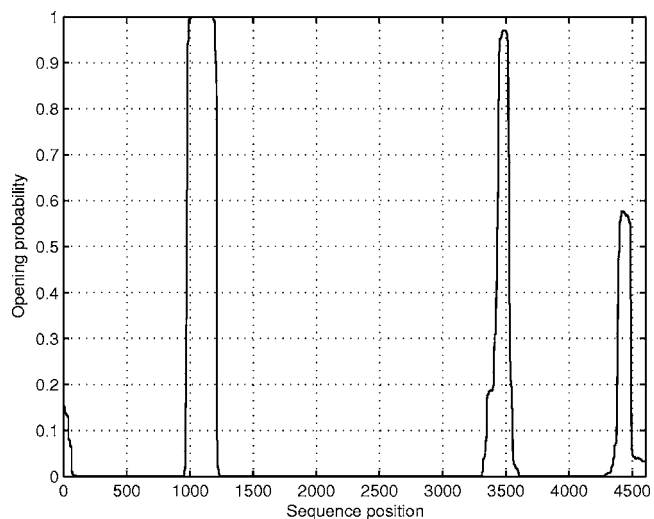


FIG. 1. PN/MCS13 opening probability at $T=310 \text{ K}$ and $\Gamma=-0.042 \text{ eV/rad}$.

Sec. II A). To do this correctly, we need to know the correlation length, or more precisely the length at which the correlation between a base pair and the rest of the sequence vanishes. Hence for a fixed base pair n we compute, using (7),

$$C_m^n = \langle r_n r_m \rangle - \langle r_n \rangle \langle r_m \rangle.$$

Figure 2 shows the correlation function C_m^n for two different values of n , $n=3289$ in the middle of the second opened bubble (see Fig. 1), and $n=2200$ in the largest closed region. Clearly, the correlation is much larger in the denatured region, but even here it does not extend beyond a few 100 base pairs.

Due to the inhomogeneity of base-pair bonding and stacking energies, DNA melting is a stepwise process with different domains melting at different temperatures. This can be

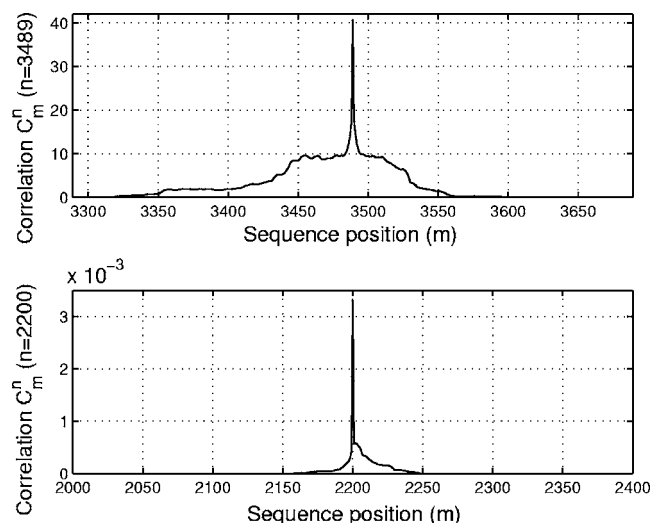


FIG. 2. Correlation function C_m^n for $n=3489$ (top) and $n=2200$ (bottom) for the PN/MCS13 sequence at $T=310 \text{ K}$ and $\Gamma=-0.042 \text{ eV/rad}$.

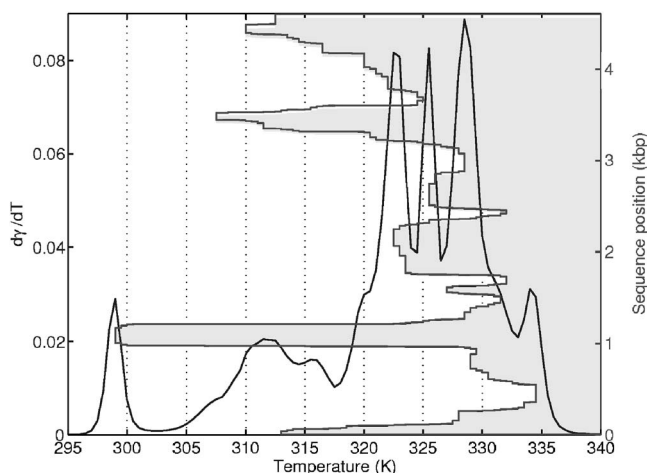


FIG. 3. PN/MCS13 differential melting curve and melting map (shaded area) (temperature increment 0.5 K, $\Gamma = -0.042$ eV/rad).

visualized by computing differential melting curves and melting maps. Let γ be the fraction of open base pairs, $\gamma = (\sum_n p_n) / N$. A differential melting curve is a plot of $d\gamma/dT$ vs temperature T . A melting map is obtained by displaying for each temperature the base pairs which have an opening probability greater than 1/2 (shaded area). Such a map gives another picture of thermodynamically stable (high melting temperature) vs unstable (low melting temperature) regions in the particular sequence.

Figure 3 shows the differential melting curve [obtained by differentiating a cubic splines interpolation of the computed values $\gamma(T)$] and the melting map for the PN/MCS13 sequence under the same condition $\Gamma = -0.042$ as in Figs. 1 and 2. We can clearly identify again the AT-rich inserted region around position 1000 which melts first, as well as the two unstable regions around positions 3500 and 4500.

Comparison of the differential melting curve and melting map with the MELTSIM result ([31] Fig. 3) shows first of all that the general shape of the melting curve is correct, but the temperature range from a completely closed to a completely denatured molecule is about twice as large in the helicoidal Peyrard-Bishop model with the current set of parameters. On the other hand, the melting map as a map depicting the successive melting order of different regions is in precise agreement with the MELTSIM melting map.

A more systematic determination of the physical value of the various energy parameters in the helicoidal model is desirable, but since the whole process of fitting model computations to experimental results of DNA denaturation in solution is quite subtle (the experimental results also depend on external conditions like, e.g., the solvent salt concentration [1,29]), it falls beyond the scope of this paper. It should also be pointed out that while such fitting was important in the early stages of the theoretical study of DNA melting, present day problems concern more the identification of stable and unstable regions and linking those to genomic content. As long as the relative strength of the various energy terms is kept within certain limits, this identification is unaffected by changing the model parameters.

In Figs. 4 and 5 we illustrate some of the effects of chang-

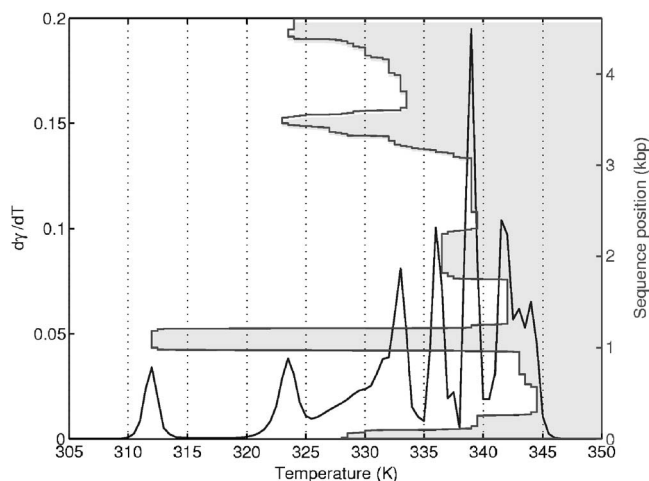


FIG. 4. PN/MCS13 differential melting curve and melting map (shaded area) with homogeneous stacking and twist energy ($K=0.1486$ eV, $E=0.0942$ eV, $\theta_0=34.81^\circ$) (temperature increment 0.5 K, $\Gamma = -0.042$ eV/rad).

ing the model parameters. Figure 4 shows the differential melting curve and melting map with homogeneous stacking and twist energy terms, where the values of K , E , and θ_0 are the averages of the values given in the Appendix. The overall identification of stable vs unstable regions remains intact, but comparison with Fig. 3 shows that considerable detail in the melting map is lost, with larger regions melting at once.

Figure 5 shows the effect of changing the relative strength of the stacking and twisting energy terms. Again they are taken homogeneous, but now with the original values of Barbi *et al.* [23]. Although with these values the transition temperature interval is of the right magnitude, the differential melting curve and melting map clearly display insufficient detail. Most notably, the two distinct unstable regions around positions 3500 and 4500 are merged into one large region.

So far, we have shown results for a chosen value $\Gamma = -0.042$ for easy comparison between the different figures,

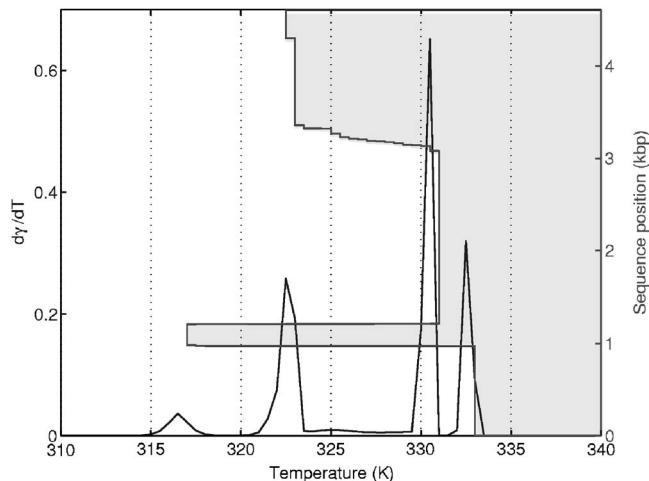


FIG. 5. PN/MCS13 differential melting curve and melting map (shaded area) with homogeneous stacking and twist energy ($K=0.65$ eV, $E=0.04$ eV, $\theta_0=34.78^\circ$) (temperature increment 0.5 K, $\Gamma = -0.042$ eV/rad).

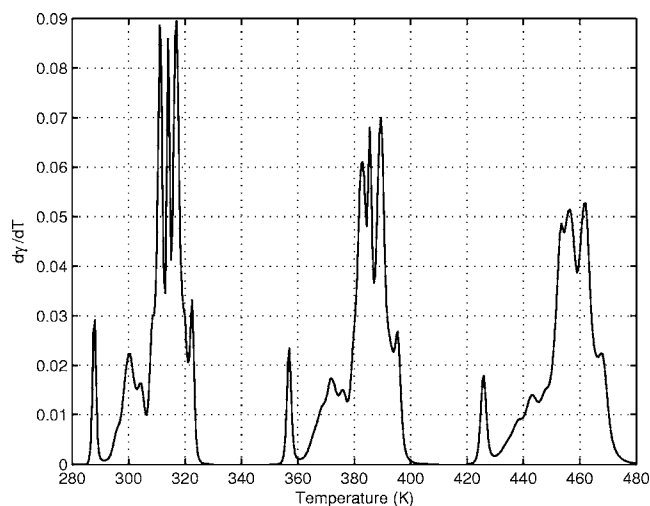


FIG. 6. PN/MCS13 differential melting curves for $\Gamma = -0.05, 0.0, \text{ and } 0.05$ eV/rad (left to right, temperature increment 0.5 K).

but other values can be considered as well. At fixed temperature, increasing Γ decreases the fraction of open base pairs, corresponding to an increase in the phase transition temperature in the thermodynamic limit [23]. What is perhaps more interesting is the fact that increasing Γ also smooths the differential melting curve (see Fig. 6), and broadens the transition; decreasing Γ has the opposite effect. Heuristically, increasing Γ effectively increases the stiffness of the double stranded DNA, which is indeed known to broaden the transition [7,34]. The value $\Gamma = 0$ plays no special role in this respect. In contrast, a recent model [35] which adds angular degrees of freedom to the Poland-Scheraga helix-coil model singles out the value $\Gamma = 0$ as special and predicts a broadening of the transition for $\Gamma < 0$ as well as $\Gamma > 0$.

Finally, we have also tested the performance of the transfer matrix algorithm on larger sequences, up to about $N \approx 3 \times 10^5$. The algorithm was written in MATLAB and run on a 2.8 GHz PC, and computed times follow the line $t = 10^{-4}N + 0.40$, with t in seconds. Comparison with [8] shows that our algorithm performs as fast as the fastest available helix-coil algorithm.

B. Linking number ensemble

In this section, we illustrate the solution of the fixed linking number ensemble, and compare it to the fixed torque ensemble as well as the helix-coil SIDD model [13], by showing example results for the C-MYC sequence ($N = 3200$), available as Example 3 on the WebSIDD server [16]. Again, the qualitative conclusions drawn from this example are valid in general. Following the outline of Sec. II B, we start by showing in Fig. 7 a plot of $F_{iq}(\Gamma) + \alpha\Gamma$ for different values of the superhelical density $\sigma = -0.06, -0.045, -0.03, \text{ and } -0.015$, corresponding to values $\alpha = (1 + \sigma)Lk_0/N = 0.572, 0.581, 0.590, \text{ and } 0.600$. As σ goes to 0, the graph becomes constant for Γ smaller than a critical value corresponding to the torque induced melting transition observed in the homogeneous model [22,23]. For nonzero σ the graph has a maximum at some value $\Gamma_0(\sigma)$

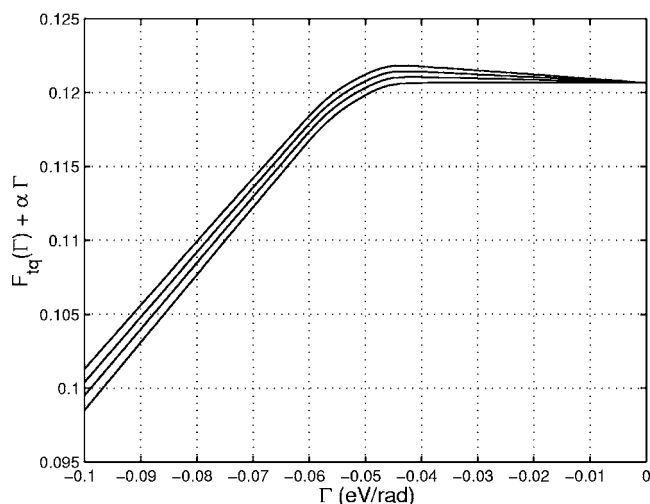


FIG. 7. C-MYC $F_{iq}(\Gamma) + \alpha\Gamma$ for $\alpha = 0.572, 0.581, 0.590, \text{ and } 0.600$ (top to bottom) at $T = 310$ K.

and this is the value we need for constructing the integration contour and for comparing the linking number and torque ensembles. We emphasize here that obtaining a very precise value of the location of the maximum is not necessary. Indeed, the integration in Eqs. (9) and (10) can be carried out along any line parallel to the imaginary axis. Taking a line at or close to the maximum will simply ensure that the function to be integrated falls off very quickly along this line.

Next we turn our attention to the integrand in Eq. (9),

$$u(\omega) = e^{-\beta N[F_{iq}(\Gamma_0 + i\omega) - F_{iq}(\Gamma_0)]} e^{-i\beta N \alpha \omega}. \quad (12)$$

Figure 8 shows the absolute value of u in a neighborhood of $\omega = 0$ for a superhelical density $\sigma = -0.03$ and temperature $T = 310$ K. For this value of σ and T , the critical point is given by $\Gamma_0 = -0.04149$. As expected, the function decays to 0 rapidly, but clearly not rapidly enough to apply a stationary phase approximation [$(\beta N)^{-1/2} = 0.003$]. Figure 9 shows the real part of u , which is the function to be integrated to obtain

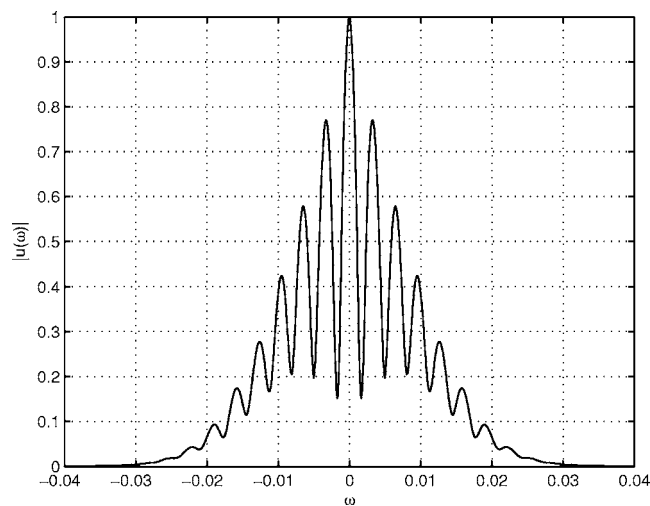


FIG. 8. C-MYC absolute value of $u(\omega)$ [Eq. (12)] at $\sigma = -0.03$ ($\alpha = 0.59$) and $T = 310$ K (ω -interval $5 \cdot 10^{-4}$).

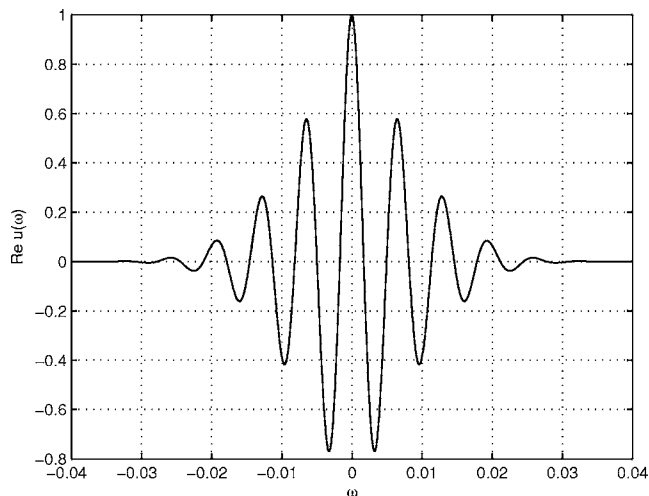


FIG. 9. C-MYC real value of $u(\omega)$ [Eq. (12)] at $\sigma = -0.03$ ($\alpha = 0.59$) and $T = 310$ K (ω -interval $5 \cdot 10^{-4}$).

the partition function in Eq. (9). Both Figs. 8 and 9 are generated by interpolating between a number of computed data points. Due to the oscillations, it is important to compute enough data points, we used an interval of $\Delta\omega = 5 \cdot 10^{-4}$. One way to determine the accuracy of the numerical approximation is to check if the expectation value of the superhelical density matches the imposed value. We find $[(2\pi)^{-1} \sum_n \langle \theta_n \rangle / Lk_0 - Lk_0] / Lk_0 = -0.03005$ which compares well with the exact value of -0.03 . Similarly, we can check how well the value of Γ_0 was determined by computing the expected helical density in the fixed torque ensemble with torque Γ_0 , and find a value of -0.03026 . This is the value obtained by differentiating the splines approximation of Fig. 7, and due to the broad maximum, better accuracy should not be expected. However, it is clear from the fast decay of the function $|u(\omega)|$ in Fig. 8, that this value of Γ_0 is accurate enough for computing expectation values in the linking number ensemble. If a more accurate value is desired, such as in Fig. 11 below, we can start from this approximate Γ_0 , and fine tune it by computing $\langle \sum_n \theta_n \rangle_{\Gamma, \Gamma}$ for some values $\Gamma \approx \Gamma_0$ until the correct linking number is found.

To visualize how the linking number constraint affects the melting behavior of a sequence by introducing an effective long range base pair coupling in the partition function, we follow Benham and Bi [17], and compare the original C-MYC sequence to a modified sequence which differs from the C-MYC sequence in a tiny fragment only. More precisely, we compute the opening probability for the C-MYC sequence at a fixed linking number ($\sigma = -0.03$) (Fig. 10, top panel), then remove from the sequence a small 44 bp segment in the center of the main untwisted region (sequence positions 781–824), and compute the opening probability for this modified sequence at the same superhelical density (Fig. 10, bottom panel).

For the C-MYC sequence, we find that there are two locations that are preferentially opened, a first, large one, between positions 760–850, and a second, smaller one, between position 2900–2950, with a much higher opening probability for the largest region. In agreement with the

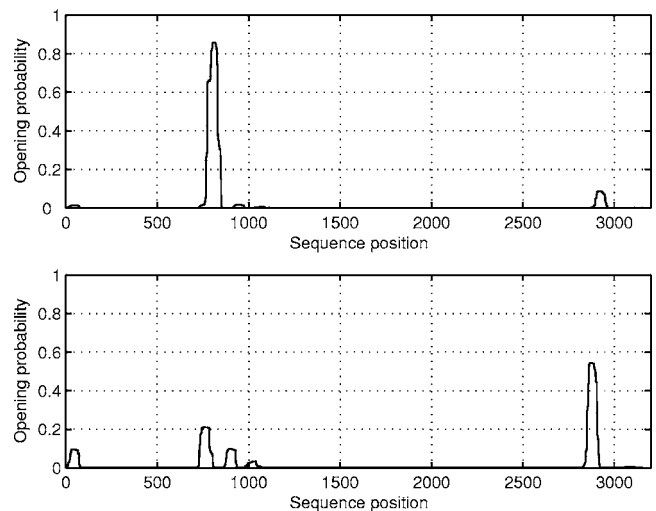


FIG. 10. The opening probability at $T = 310$ K and fixed superhelical density $\sigma = -0.03$ for the C-MYC sequence ($N = 3200$) (top) and the modified C-MYC sequence ($N = 3156$) (bottom).

SIDD-model [17] we see that a small modification of the sequence is sufficient to shift the main opening activity to the second region.

In Fig. 11, we show the corresponding opening probabilities in the fixed torque ensemble. For a fair comparison, we adjusted the torque values for both sequences separately to return a superhelical density expectation value $\sigma = -0.03$ with high accuracy. As a final check that we are comparing both ensembles with the right parameters, we compute the total fraction of open base pairs $N^{-1} \sum_n p_n$, and find that it is equal to 0.018 for both top panels of Figs. 10 and 11, and equal to 0.017 for both bottom panels.

The main qualitative difference that can be observed between both ensembles is that the effect of removing the small

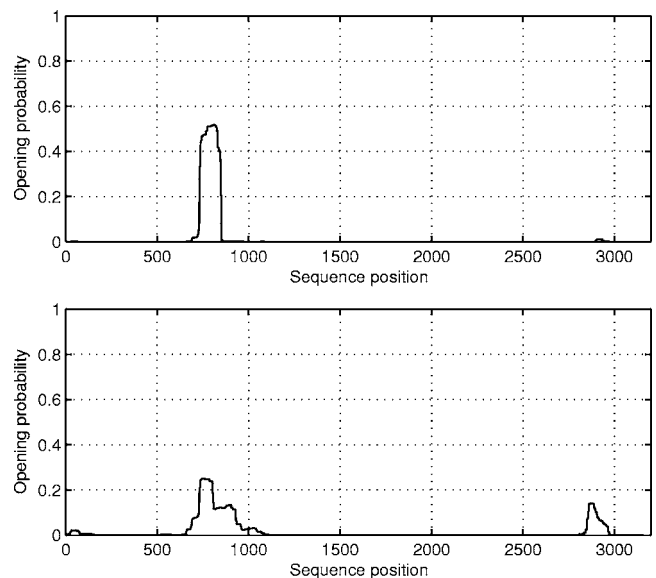


FIG. 11. The opening probability at $T = 310$ K in the fixed torque ensemble for the C-MYC sequence ($N = 3200$, $\Gamma = -0.041475$, $\langle \sigma \rangle_{\Gamma} = -0.03004$) (top) and the modified C-MYC sequence ($N = 3156$, $\Gamma = -0.044365$, $\langle \sigma \rangle_{\Gamma} = -0.03003$) (bottom).

segment is much more localized in the torque ensemble, and the first region is still dominant. If we consider a base pair to be open if $p_n > 0.5$, like in constructing the melting maps in Sec. III A, we see that in the linking number ensemble the open region shifts from the left to the right upon modifying the sequence, while in the torque ensemble, the open region disappears.

IV. CONCLUSIONS AND OUTLOOK

In this paper we have connected the particle-lattice helicoidal Peyrard-Bishop model to more familiar Ising-type models for inhomogeneous DNA melting. In the simplest setting of a fixed external torque, the model has the same melting behavior as the Poland-Scheraga helix-coil model. Since the numerical integrations needed to compute the profiles in the particle-lattice model can be carried out using a limited number of discretization points, and since the interactions are only nearest neighbor, we have obtained a method to compute melting profiles which is both very simple to implement and very efficient to execute, and which is therefore highly attractive to analyze very long, or even whole genome sequences.

Furthermore we have shown that also the more complicated setting of a fixed linking number can be treated and that the results are in agreement with Benham's SIDD model. The algorithm is again simple to implement and consists of numerically integrating the fixed torque results over a small range of complex torque values. Some of the points raised here, such as the inequivalence of ensembles, are worthwhile of investigating mathematically more rigorously in the setting of the homogeneous helicoidal Peyrard-Bishop model to see if they persist in the thermodynamic limit.

The equivalence between nearest-neighbor lattice models with continuous degrees of freedom on the one hand, and Ising models with loop entropy weights or long-range interaction on the other hand, raises more fundamental questions as well. A better understanding of this equivalence will presumably lead to a better understanding of nonlinear phenomena in one-dimensional systems.

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APPENDIX: ENERGY PARAMETERS

In this appendix we collect the various parameters used in the potential energy (1). All lengths are measured in Å, energies in eV, and angles in rad.

For the depth D_i of the Morse potentials, we choose values close to the value 0.15 of [23], but taking into account that a C-G base pair has a 1.5 times stronger bond than an A-T base pair. For the widths a_i we take the values of [29].

$$D_1 = D_4 = 0.12, \quad D_2 = D_3 = 0.18,$$

$$a_1 = a_4 = 4.2 \quad a_2 = a_3 = 6.9.$$

The equilibrium distance r_0 is equal to 10.

The length $\ell_{n,n+1}$ between successive nucleotides on the same DNA strand in the twist energy term is given by

$$\ell_{n,n+1} = \sqrt{h^2 + r_n^2 + r_{n+1}^2 - 2r_n r_{n+1} \cos \theta_n}, \quad (19)$$

where $h=3.4$ is the fixed vertical distance between base pairs. The rest length $\ell_{n,n+1}^{(0)}$ is step dependent and given by

$$\ell_{s_n s_{n+1}}^{(0)} = \sqrt{h^2 + 4r_0^2 \sin^2 \left(\frac{1}{2} \theta_{s_n s_{n+1}}^{(0)} \right)},$$

where $\theta_{s_n s_{n+1}}^{(0)}$ is the average helical twist angle of the given step, taken from the database of El Hassan and Calladine [30]

$$\theta^{(0)} = \frac{2\pi}{360} \times \begin{pmatrix} 35.9 & 32.9 & 34.8 & 32.4 \\ 37.4 & 31.9 & 35.1 & 34.8 \\ 37.8 & 37.4 & 31.9 & 32.9 \\ 30.6 & 37.8 & 37.4 & 35.9 \end{pmatrix}.$$

The parameter E is taken inversely proportional to the twist angle standard deviations, taken from the same database [30],

$$E = 0.4 \times \begin{pmatrix} 0.3030 & 0.2632 & 0.2083 & 0.3571 \\ 0.1053 & 0.2703 & 0.1887 & 0.2083 \\ 0.2632 & 0.2500 & 0.2703 & 0.2632 \\ 0.1493 & 0.2632 & 0.1053 & 0.3030 \end{pmatrix}.$$

Similarly, the stacking energy parameter K is taken inversely proportional to the slide standard deviations of [30]:

$$K = 0.1 \times \begin{pmatrix} 3.5714 & 1.4085 & 1.2195 & 2.0833 \\ 0.8130 & 0.8547 & 0.9804 & 1.2195 \\ 1.4493 & 1.1628 & 0.8547 & 1.4085 \\ 0.9174 & 1.4493 & 0.8130 & 3.5714 \end{pmatrix}.$$

The constant α in the exponential is put equal to 0.5 as in [23].

- [1] D. Poland and H. Scheraga, *Theory of Helix-Coil Transitions in Biopolymers* (Academic, New York, 1970).
 [2] R. Wartell and A. Benight, *Phys. Rep.* **126**, 67 (1985).
 [3] D. Poland, *Biopolymers* **13**, 1859 (1974).

- [4] M. Fixman and J. Freire, *Biopolymers* **16**, 2693 (1977).
 [5] E. Yeramian, *Gene* **255**, 139 (2000).
 [6] E. Carlon, M. Malki, and R. Blossey, *Phys. Rev. Lett.* **94**, 178101 (2005).

- [7] R. Blossey and E. Carlon, *Phys. Rev. E* **68**, 061911 (2003).
- [8] E. Tøstesen, F. Liu, T. Jenssen, and E. Hovig, *Biopolymers* **70**, 364 (2003).
- [9] T. Garel and H. Orland, e-print q-bio/0402037.
- [10] H. Kramers and G. Wannier, *Phys. Rev.* **60**, 252 (1941).
- [11] L. Onsager, *Phys. Rev.* **65**, 117 (1944).
- [12] D. Poland, *Biopolymers* **73**, 216 (2004).
- [13] R. M. Fye and C. J. Benham, *Phys. Rev. E* **59**, 3408 (1999).
- [14] C. Benham, *Proc. Natl. Acad. Sci. U.S.A.* **90**, 2999 (1993).
- [15] C. Benham, *J. Mol. Biol.* **255**, 425 (1996).
- [16] C.-P. Bi and C. Benham, *Bioinformatics* **20**, 1477 (2004); <http://www.genomecenter.ucdavis.edu/benham/sidd/>
- [17] C. Benham and C.-P. Bi, *J. Comput. Biol.* **11**, 519 (2004).
- [18] M. Peyrard and A. R. Bishop, *Phys. Rev. Lett.* **62**, 2755 (1989).
- [19] M. Peyrard, *Nonlinearity* **17**, R1 (2004).
- [20] T. Dauxois, M. Peyrard, and A. R. Bishop, *Phys. Rev. E* **47**, R44 (1993).
- [21] M. Barbi, S. Cocco, and M. Peyrard, *Phys. Lett. A* **253**, 358 (1999).
- [22] S. Cocco and R. Monasson, *Phys. Rev. Lett.* **83**, 5178 (1999).
- [23] M. Barbi, S. Lepri, M. Peyrard, and N. Theodorakopoulos, *Phys. Rev. E* **68**, 061909 (2003).
- [24] D. Cule and T. Hwa, *Phys. Rev. Lett.* **79**, 2375 (1997).
- [25] Y. L. Zhang, W. M. Zheng, J. X. Liu, and Y. Z. Chen, *Phys. Rev. E* **56**, 7100 (1997).
- [26] T. Strick, M.-N. Dessinges, G. Charvin, N. Dekker, J.-F. Allemand, D. Bensimon, and V. Croquette, *Rep. Prog. Phys.* **66**, 1 (2003).
- [27] H. Zhou, Z. Yang, and Z. c. Ou-Yang, *Phys. Rev. Lett.* **82**, 4560 (1999).
- [28] H. Zhou, Z. Yang, and Z. c. Ou-Yang, *Phys. Rev. E* **62**, 1045 (2000).
- [29] A. Campa and A. Giansanti, *Phys. Rev. E* **58**, 3585 (1998).
- [30] M. El Hassan and C. Calladine, *Philos. Trans. R. Soc. London, Ser. A* **355**, 43 (1997).
- [31] R. Blake, J. W. Bizarro, J. Blake, G. Day, S. Delcourt, J. Knowles, and J. J. SantaLucia, *Bioinformatics* **15**, 370 (1999).
- [32] W. Press, S. Teukolsky, W. Vetterling, and B. Flannery, *Numerical Recipes in C* (Cambridge University Press, Cambridge, 1992).
- [33] R. Blake and S. Delcourt, *Nucleic Acids Res.* **26**, 3323 (1998).
- [34] E. Carlon, E. Orlandini, and A. L. Stella, *Phys. Rev. Lett.* **88**, 198101 (2002).
- [35] T. Garel, H. Orland, and E. Yeramian, e-print q-bio/0407036.
- [36] GenBank/EMBL accession number J01749. <http://www.ebi.ac.uk/embl/>